



A Concise Review -Versatility of Amniotic Epithelial Stem Cells (hAECs)

Author

D.Hettiaracchchi¹

¹Human Genetics Unit, Faculty of Medicine, University of Colombo, Sri Lanka

Abstract

Amniotic epithelial cells are generated from amnioblasts on the eighth day after fertilization and constitute the inner layer of the amnion. Many studies have demonstrated the differentiation potential of human amniotic epithelial cells (HAEC) into cells of all three germ layers. Its easy accessibility without any ethical constraints has made it an ideal non-controversial source of primary cells that can be differentiated in a plethora of organ specific lineages. The aim of this review is to highlight the versatility of hAECs whilst exploring its tri-lineage differentiation potential. Which makes these cells an ideal candidate for disease modeling and cellular replacement therapy.

Keywords: *Amniotic epithelial cells, cellular replacement therapy, disease modeling.*

Introduction

Human amniotic membrane is a rich source of stem cells. Isolated amniotic epithelial cells (hAECs) have especial properties that make them truly unique in comparison to other stem cell populations. Its ability to differentiate towards all three-germ layers make hAECs pluripotent similar to embryonic stem cells (ESC) without the ethical constraints since placenta are discarded following delivery. In addition hAECs are non-tumorigenic and have immunosuppressive properties⁽¹⁻³⁾. Due to these favorable characteristics of hAECs they have a variety of applications in cellular replacement therapies in diseases such as diabetes, neurodegenerative disorders, ischemic diseases (myocardial and brain infarct), spinal cord injury, etc. They can also be employed as a platform for drug and toxicology screening⁽⁴⁻⁶⁾. Through this literature review I wish to highlight the differentiation potential of hAECs along with their applications in modern medicine.

Practical advantages and differentiation potential of hAECs

Amniotic epithelial stem cells can be isolated with ease from the amniotic membrane following a 4-quadrant blunt dissection method. Followed by stripping off the amniotic membrane from the chorion and digested with trypsin or other digestive enzymes⁽⁷⁻⁹⁾. Epithelial cells are usually released following a brief digestive period of 20-30 minutes. These cells have a plastic adherence capacity and proliferate robustly in a simple complete media (Dulbecco's modified Eagle's medium supplemented with 5%–10% serum and L- Glutamine). They can be subcultured up to 5-6 passages before proliferation ceases and display typical cuboidal epithelial morphology^(10,11). They express a variety of cell surface antigens including ATP-binding cassette transporter G2 (ABCG2/BCRP), CD9, CD24, E-cadherin, integrins $\alpha 6$ and $\beta 1$, c-met (hepatocyte growth factor receptor), stage-specific embryonic antigens

(SSEAs) 3 and 4, and tumor rejection antigens 1-60 and 1-81, their overall low antigenicity makes them ideal candidates for cellular therapies^(11,12). Additionally they express molecular markers of pluripotent stem cells including octamer-binding protein 4 (OCT-4), SRY-related HMG-box gene 2 (SOX-2), and Nanog⁽⁸⁾.

Ectodermal Derivatives

The ability of hAECs to differentiate to neural cells was demonstrated by Sakuragawa and coworkers in 1996⁽¹³⁾. Studies conducted in animal models have shown that hAECs can promote neural cell differentiation, reduce secondary neural damage associated with inflammation and apoptosis in spinal cord injury^(14,15). It can also promote re-myelination and sprouting of nerve fibers whilst improving functional recovery⁽¹⁶⁻²⁰⁾. Their ability to synthesize catecholamines including dopamine (DA) has explored the possibility of hAEC serving as a donor for transplantation therapy of Parkinson's disease (PD)⁽²¹⁾. Furthermore researches have shown that hAEC significantly suppressed splenocyte proliferation *in vitro* and has the potential to attenuate multiple sclerosis (MS) in mouse models⁽²²⁾.

Endodermal Derivatives

Studies have demonstrated expression of hepatocyte-related genes in hAEC such as albumin, α 1-antitrypsin etc. Cultivated hAEC has also demonstrated albumin production, glycogen storage, and albumin secretion consistent with the hepatocyte gene expression profile⁽²³⁾. Hence hAEC represent a noncontroversial source of cells for liver-based regenerative medicine^(24,25). hAEC can also be induced to differentiate into pancreatic β -cells. It has been demonstrated that multiple such genes including insulin, pancreas duodenum homeobox-1, paired box gene 6, NK2 transcription factor-related locus 2, Islet 1, glucokinase, and glucose transporter-2 are expressed in these differentiated progenitors. Scientists have also discovered that

these cells release C-peptide in a glucose-regulated manner in response to extracellular stimuli. These findings indicated that hAEC might be a new source for cell replacement therapy in type 1 diabetes^(26,27).

Mesodermal Derivatives

It has been shown that freshly isolated hAEC express cardiac-specific transcription factor GATA4; cardiac-specific genes such as myosin light chain (MLC)-2a, MLC-2v, cTnI, and cTnT; and the α -subunits of the cardiac-specific L-type calcium channel (α 1c) and the transient outward potassium channel (Kv4.3). Upon stimulation with basic fibroblast growth factor (bFGF) or activin A these cells have shown to express Nkx2.5, a specific transcription factor for cardiomyocytes, and a cardiac-specific marker, atrial natriuretic peptide (ANP). Co-culture experiments have confirmed that hAEC cells are able to both integrate into cardiac tissues and differentiate into cardiomyocyte-like cells⁽²⁸⁻³¹⁾. hAEC also express chondrocyte related genes, including SOX-9, SOX-5, SOX-6, bone morphogenetic proteins (BMP)-2 and -4, as well as BMP receptors. Recent studies have shown that implanted hAECs aid in cartilage repair in porcine models⁽³²⁾. One study has demonstrated that *in vitro* induction of viable human amnion expresses cartilage-specific markers and accumulates GAGs within the biomatrix⁽³³⁾.

Conclusion

Owing to its tri-lineage differentiation potential hAEC are a versatile cell population that can be easily harvested and propagated into a desired tissue type or a disease model.

Reference

1. Antoniadou, Eleni, and Anna L. David. "Placental stem cells." *Best Practice & Research Clinical Obstetrics & Gynaecology* 31 (2016): 13-29.
2. Matikainen, Tiina, and Jarmo Laine. "Placenta—an alternative source of stem

- cells." *Toxicology and applied pharmacology* 207.2 (2005): 544-549.
3. Maymó, Julieta L., et al. "Proliferation and survival of human amniotic epithelial cells during their hepatic differentiation." *PloS one* 13.1 (2018): e0191489.
 4. Davila, J.C., Cezar, G.G., Thiede, M., Strom, S., Miki, T., Trosko, J., 2004. Use and application of stem cells in toxicology. *Toxicol. Sci.* 79, 214 – 223.
 5. Gorba, T., Allsopp, T.E., 2003. Pharmacological potential of embryonic stem cells. *Pharmacol. Res.* 47, 269–278.
 6. Rolletschek, A., Blyszczuk, P., Wobus, A.M., 2004. Embryonic stem-cell derived cardiac, neuronal and pancreatic cells as model systems to study toxicological effects. *Toxicol. Lett.* 149, 361–369.
 7. Terada, Satoshi, et al. "Inducing proliferation of human amniotic epithelial (HAE) cells for cell therapy." *Cell transplantation* 9.5 (2000): 701-704.
 8. Miki, Toshio, et al. "Stem cell characteristics of amniotic epithelial cells." *Stem cells* 23.10 (2005): 1549-1559.
 9. Miki, Toshio, et al. "Isolation of amniotic epithelial stem cells." *Current protocols in stem cell biology* (2010): 1E-3.
 10. Ochsenbein-Kölble, Nicole, et al. "Inducing proliferation of human amnion epithelial and mesenchymal cells for prospective engineering of membrane repair." *Journal of perinatal medicine* 31.4 (2003): 287-294.
 11. Parolini, Ornella, et al. "Concise review: isolation and characterization of cells from human term placenta: outcome of the first international Workshop on Placenta Derived Stem Cells." *Stem cells* 26.2 (2008): 300-311.
 12. Miki, Toshio, and Stephen C. Strom. "Amnion-derived pluripotent/multipotent stem cells." *Stem cell reviews* 2.2 (2006): 133-141.
 13. Sakuragawa, Norio, et al. "Expression of markers for both neuronal and glial cells in human amniotic epithelial cells." *Neuroscience letters* 209.1 (1996): 9-
 14. Gao, Yuhua, et al. "Combination of melatonin and Wnt- 4 promotes neural cell differentiation in bovine amniotic epithelial cells and recovery from spinal cord injury." *Journal of pineal research* 60.3 (2016): 303-312.
 15. Gao S, Ding J, Xiao HJ, et al. Anti-inflammatory and anti- apoptotic effect of combined treatment with methylprednisolone and amniotic membrane mesenchymal stem cells after spinal cord injury in rats. *Neurochem Res.* 2014;39:1544-1552.
 16. Roh DH, Seo MS, Choi HS, et al. Transplantation of human umbilical cord blood or amniotic epithelial stem cells alleviates mechanical allodynia after spinal cord injury in rats. *Cell Trans- plant.* 2013;22:1577-1590.
 17. Xue H, Zhang XY, Liu JM, Song Y, Li YF, Chen D. Development of a chemically extracted acellular muscle scaffold seeded with amniotic epithelial cells to promote spinal cord repair. *J Biomed Mater Res A.* 2013;101:145-156.
 18. Meng XT, Li C, Dong ZY, et al. Co-transplantation of bFGF- expressing amniotic epithelial cells and neural stem cells pro- motes functional recovery in spinal cord-injured rats. *Cell Biol Int.* 2008;32:1546-1558.
 19. Wu ZY, Hui GZ, Lu Y, Wu X, Guo LH. Transplantation of human amniotic epithelial cells improves hindlimb function in rats with spinal cord injury. *Chin Med J (Engl).* 2006;119: 2101-2107.
 20. Shaw, K. Aaron, et al. "The Science and Clinical Applications of Placental Tissues in Spine Surgery." *Global Spine Journal* (2018): 2192568217747573.

21. Kakishita, Koji, et al. "Human amniotic epithelial cells produce dopamine and survive after implantation into the striatum of a rat model of Parkinson's disease: a potential source of donor for transplantation therapy." *Experimental neurology* 165.1 (2000): 27-34.
22. Liu, Yu Han, et al. "Amniotic epithelial cells from the human placenta potently suppress a mouse model of multiple sclerosis." *PloS one* 7.4 (2012): e35758.
23. Takashima, Seiji, et al. "Human amniotic epithelial cells possess hepatocyte-like characteristics and functions." *Cell structure and function* 29.3 (2004): 73-84.
24. Marongiu, Fabio, et al. "Hepatic differentiation of amniotic epithelial cells." *Hepatology* 53.5 (2011): 1719-1729.
25. Miki, Toshio, et al. "Production of hepatocyte-like cells from human amnion." *Hepatocyte Transplantation*. Humana Press, Totowa, NJ, 2009. 155-168.
26. Hou, Yanan, et al. "Human amnion epithelial cells can be induced to differentiate into functional insulin producing cells." *Acta biochimica et biophysica Sinica* 40.9 (2008): 830-839.
27. Miki T, Lehmann T, Cai H, Stolz DB, Strom SC. Stem cell characteristics of amniotic epithelial cells. *Stem Cells* 2005, 23: 1549–1559
28. Zhao P, Ise H, Hongo M, Ota M, Konishi I, Nikaido T. Human amniotic mesenchymal cells have some characteristics of cardiomyocytes. *Transplantation*. 2005;79:528–535.
29. Tanaka M, Chen Z, Bartunkova S, Yamasaki N, Izumo S. The cardiac homeobox gene *Csx / Nkx2.5* lies genetically upstream of multiple genes essential for heart development. *Development*. 1999;126:1269–1280.
30. Kasahara H, Usheva A, Ueyama T, Aoki H, Horikoshi N, Izumo S. Characterization of homo- and heterodimerization of cardiac *Csx / Nkx2.5* homeoprotein. *J Biol Chem*. 2001;276:4570–4580.
31. Logan M, Mohun T. Induction of cardiac muscle differentiation in isolated animal pole explants of *Xenopus laevis* embryos. *Development* 1993;118:865–875.
32. Muiños-López, E., et al. "An In vitro porcine study of repairing articular cartilage with human amniotic membrane epithelial and mesenchymal stem cells." *Osteoarthritis and Cartilage* 20 (2012): S273.
33. Lindenmair, Andrea, et al. "Intact human amniotic membrane differentiated towards the chondrogenic lineage." *Cell and tissue banking* 15.2 (2014): 213-225.