



Bacterial Isolates in ICU of A Tertiary Care Hospital in North East India: A Three Year Experience

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Abstract

Introduction: *The incidence of nosocomial infections in ICU is 4-5 times greater than in general ward. The study was to detect the pattern of bacterial isolates in ICU.*

Methods: *Specimens collected from respiratory tract, urine, blood and wound. The specimens collected sent to laboratory to identify the isolates.*

Result: *Respiratory tract infections were most prevalent with 79.5% followed by urinary tract (11.1%), blood born infection (8.1) and wound infection 1.2%. Gram negative infections like klebsiella, being the most frequent species with 54.6% followed by E coli and pseudomonas 15.8% and 14.2% respectively.*

Conclusion: *Gram negative infections were the predominant cause of icu acquired infection in the region of North East India. Pattern and prevalence of bacterial isolates in icu tends to change with time.*

Keywords: *nosocomial infections, ICU, Gram negative infections, pattern changes.*

Introduction

The incidence of nosocomial infections in ICU is 4-5 times greater than in general ward⁽¹⁾. Critically ill patients are always at higher risk of developing nosocomial infections with resistant strains. Klebsiella, Pseudomonas, Acinetobacter, methicillin resistant Staphylococcus aureus (MRSA), vancomycin-resistant Enterococcus (VRE), Clostridium difficile, extended-spectrum β -lactamase-producing GNB (ESBL) etc. Infections caused by these microorganisms increase hospital stay and attributes to high mortality, morbidity, financial burden^(2, 3). The study was conducted to detect the pattern of

bacterial isolates in ICU of a tertiary care hospital in north east India.

Materials and Methods

The present study was an observational retrospective data analysis on bacterial isolates and changing pattern of icu acquired infections. Study was conducted in Emergency ICU of Emergency Medicine Department. Gauhati Medical College and Hospital, Guwahati for three years from November 2013 to October 2016 and divided into three phases each comprising of twelve months. Phase 1: Nov 2013- Oct 2014 Phase 2: Nov 2014- Oct 2015 Phase 3: Nov 2015- Oct 2016 March 2003 to February

2004. The study proposal was reviewed and approved by the institutional ethics committee of the hospital.

Selection of cases: Inclusion Criteria- Patients developing signs & symptoms of infections (fever, leucocytosis, shock with suggestive of septic etiology, increased amount & purulence of tracheal secretions, hematuria, purulent surgical wounds) after staying 48hours or more in EICU with positive aerobic bacterial cultures were included in the study.

Exclusion Criteria- Febrile / signs of infection on ICU admission staying less than 48 hours in the ICU. Transferred from another ICU. Positive cultures which were thought to be Contaminations & Commensals. All positive swabs of eye, ear, nose, throat and genital cultures. Positive fungal cultures were excluded from the study.

Specimens collected were from Respiratory - Tracheal aspirate, ET tube tip, Sputum. Urine (including Foley’s catheter tips). Blood (including tip of central venous catheters). Wound swab from Surgical wound, bed sores.

The specimens for antimicrobial sensitivity testing were studied by Gram stains and culture growth on nutrient, blood and MacConkey agar to identify the isolates. Antibiotic susceptibility testing was performed by disc diffusion method (modified Kirby Bauer method) on Muller Hinton agar (For all antimicrobials tested, MIC interpretive standards were defined according to CLSI breakpoints (Clinical and Laboratory Standard Institute) but study on antibiotic susceptibility pattern not included in the study.

Results & Observations

Majority of the patients in the study were male with 66.1% with mean age of 48 years. Average icu stay of all patients were 18 days. Most of the patients were trauma (32.4%) followed by neurological patients (29.7%) in the form of stroke or other neurological disorders [table1]. Out of 1905 icu admissions in three years 1028 patients were eligible to be included in the study with inclusion and exclusion criteria and of which 138 patients came out to be culture positive. Total culture positive samples were 161 as some patients showed

positive culture from more than one site[table2]. Respiratory tract gives positive isolate most abundantly with 79.5% followed by urine 11.2% blood 8.1% and wound infection1.2%[Fig 1,table 3]. Of total isolates 7.4% were gram positive cocci, predominantly staphylococcus 5% and 92.6% were gram negative bacilli in the form of klebsiella, being the most frequent species with 54.6% followed by e coli and pseudomonas 15.8% and 14.2% respectively. Bacteriology of respiratory tract infection shows klebsiella, pseudomonas, E coli, staph aureus, acinetobacter, proteus and citobacter. Urinary tract infections showed similar bacterial isolats but more of E coli 18% and enterococcus 2%. Blood and urine also showed similar isolate more often klebsiella and E coli. [table 4,5,6,7,8]

Figure 1: pie diagram of different sites of icu acquired infections.

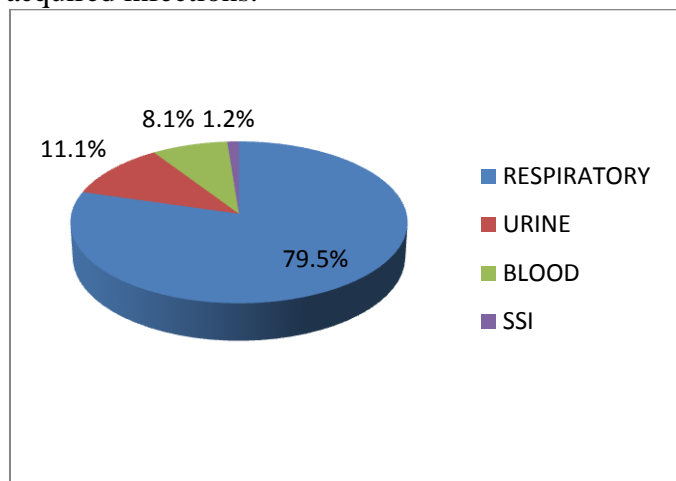


Table 1: Baseline characteristics of patients during three year period.

• Sex, male, n (%)	680 (66.1%)
• Age, years (mean)	48.6 ± 12.4
• Average length of ICU-stay, days	18.3±8.7
• Trauma	334 (32.4%)
• Neurological disorders	306 (29.7%)
• Immediate postoperative patient	95 (9.2%).
• Pulmonary diseases,	66 (6.4%)
• Diabetes mellitus,	65 (6.3%)
• Undiagnosed diseases	55 (5.3%)
• Renal diseases	46 (4.4%)
• Cardiovascular diseases	41(3.9%)
• Malignancy	20 (1.9%)
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Table 2: Total number of icu admission with culture positive patients

	Phase 1	Phase 2	Phase 3	Total
Total admitted patients	578	662	665	1905
Included in study	273	380	375	1028 (53%)
Culture positive isolates	56 (49)	48 (43)	57 (46)	161 (138) (15%)

Table 3: Types of ICU acquired Infections

SPECIMEN	Frequency	%
RESPIRATORY	128	79.5 %
URINE	18	11.1 %
BLOOD	13	8.1 %
WOUND	2	1.2 %
TOTAL	161	100 %

Table 4: Etiological agents in infections acquired in ICU

ORGANISM	Phase 1	Phase 2	Phase 3	Total	
GRAM NEGATIVE BACILLI (92.6%)	KLEBSIELLA SPP	25 (44.6%)	31 (62.5%)	32 (56.1%)	88 (54.6%)
	E COLI	12	4	10	26 (15.8%)
GRAM POSITIVE COCCI (7.4%)	PSEUDOMONAS SPP	12	4	7	23 (14.2%)
	PROTEUS SPP	1	2	2	5 (3.1%)
	ACINETOBACTER	1	1	4	6 (3.7%)
GRAM POSITIVE COCCI (7.4%)	CITROBACTER KOSERI	0	1	0	1 (0.6%)
	STAPH AUREUS	5 (MRSA =2)	1	2 (MRSA 1)	8 (5%) 3MRSA
	COAGULASE NEGATIVE STAPH	0	1	1	2 (1.2%)
	ENTEROCOCCUS	0	1	1	2 (1.2%)
TOTAL	56	48	57	161 (100%)	

Table 5: Bacteriology of respiratory tract infection.

ORGANISM	Phase 1	Phase 2	Phase 3	Total
KLEBSIELLA	23	25	28	76 (59.3%)
PSEUDOMONAS	11	4	5	20 (15.6%)
STAPH AUREUS	4 (MRSA =1)	1	2 (MRSA=1)	7 (MRSA 2) (5.4%)
E COLI	3	1	1	5 (27.7%)
ACINETOBACTER	1	1	3	5 (3.9%)
PROTEUS MIRABILIS	0	0	1	1 (0.7%)
CITROBACTER	0	1	0	1 (0.7%)
TOTAL	48	36	44	128 (100%)

Table 6: Bacteriology of urinary tract infection.

ORGANISM	Phase 1	Phase 2	Phase 3	Total
KLEB PNEUMONIAE	1	2	2	5 (27.7%)
PSEUDOMONAS SPP	1	0	2	3 (16.7%)
PROTEUS	1	1	0	2 (11.1%)
E COLI	9	2	7	18 (14%)
STAPH AUREUS	1 (MRSA)	0	0	1 (5.6%)
ENTEROCOCCUS	0	1	1	2 (11.1%)
TOTAL	7	5	6	18 (100%)

Table 7: Bacteriology of blood stream infection

ORGANISM	Phase 1	Phase 2	Phase 3	Total
KLEB PNEUMONIAE	1	3	2	6 (46.2%)
PROTEUS MIRABILIS	0	1	1	2 (15.4%)
E COLI	0	1	1	2 (15.4%)
COAGULASE NEGATIVE STAPH	0	1	1	2 (15.4%)
ACINETOBACTER	0	0	1	1 (7.6%)
TOTAL	1	6	6	13 (100%)

Table 8: Bacteriology of wound infection.

ORGANISM	Phase 1	Phase 2	Phase 3	Total
KLEBSIELLA SPP	0	1	0	1 (50%)
E COLI	0	0	1	1 (50%)
TOTAL	0	1	1	2 (100%)

Discussion

Infections acquired during stay in the ICU is very common and inevitable because of reduced host defenses, invasive devices use, damage of anatomical barrier in the form of intubation, mechanical ventilation, frequent suctioning, catheterization etc and several drugs like sedatives, muscle relaxants, H2 blockers, over use of antibiotic itself create a predisposition for infections [4,5,6].

The present study was an observational retrospective data analysis on bacterial isolates of ICU acquired infections. The study was conducted in Emergency ICU of Emergency Medicine Department for three consecutive years. Trauma and neurological patients in the form of stroke were maximum with male predominance that received in our ICU. [7,8]

Infection rate in the current study was 15.0%. The ICU infection rates may vary between centers, depending on types of patients and ICU, standard of laboratory, techniques of sample collection, diagnostic criteria etc. Current study was in single centre and for 3 consecutive year, bronchoalveolar lavage (BAL) or protected specimen brush (PSB) sample could not be collected as bronchoscope was not available. Many studies are multicentric and 1-day point-prevalence study. So percentage of acquired infections in the ICU worldwide shows wide range from 9.6 to 48.7%. [9,10,11]. In the current study, intubation and mechanical ventilation, Foleys catheter and central venous catheter, were main source of infections. Invasive procedures, mechanical ventilation, prolong ICU stay, inability to maintain proper nutrition etc have been reported as risk factors for mortality in many studies [12,13,14]

Respiratory tract infection is reported to be the most frequent site of infection in the ICU, followed by urinary tract and bloodstream infection respectively in the present study. Most of the patients were intubated or ventilated. Various studies showed pneumonia is predominant cause of ICU acquired infection. [15,16,17,18,19]

Infection with multidrug resistant pathogens develops because of frequent use of broad spectrum antibiotics and causes an increase in morbidity, mortality, and economic burden. [20,21,22,23].

Pseudomonas tends to be most resistant to treat followed by *Klebsiella*, *Acinetobacter baumannii* and *E. coli* in our experience. [24,25] The impact of increased length of ICU stay on nosocomial infections and mortality has been investigated in many studies. In the majority of the studies ICU-acquired infection was found to be an independent risk factor of increased length of ICU stay. In this study, infection with *Klebsiella pneumoniae* followed by *E. coli*, *Pseudomonas*, *Staphylococcus* and *Acinetobacter baumannii* respectively were major causative organism. Tracheal aspirate harbor most of *Klebsiella* and *Pseudomonas*. Urine sample shows most of *Klebsiella* and *E. coli*. Here in our region of North East India *Klebsiella* being most abundant isolates in ICU similar to the majority of studies in which gram-negative organisms were the predominant agent and *Staphylococcus* is still not a frequent isolate in our ICU. *Acinetobacter baumannii* culture tends to increase gradually. [26,27,28,29,30].

Conclusion

Gram negative infections were the predominant cause of ICU acquired infection in the region of North East India. Pattern and prevalence of bacterial isolates in ICU tends to change with time. Prolong ICU stay, poor nutrition, invasive techniques are few causes that increases the chance of ICU acquired infection. MDR organisms are increasing day by day. Antibiotic stewardship and strict infection control protocol particularly in ICU are need of the hour to prevent flaring up of antimicrobial resistance.

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Declaration

Conflicting Interests: The authors declare that they have no conflict of interest.

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References

1. Kollef MH, Fraser VJ. Antibiotic resistance in intensive care unit setting. *Ann Intern Med* 2001; 134: 298-314.
2. Humphreys H, Winter B, Paul M. Infections in the adult intensive care unit: Springer Science & Business Media; 2012.
3. Hosein IK, Hill DW, Jenkins LE, Magee JT. Clinical significance of emergence of bacterial resistance in the hospital environment. *Sym Ser Soc J Appl Microbiol*, 2002; 31: 90S-7S.
4. Ak O, Batirel A, Ozer S, Colakoglu S. Nosocomial infections and risk factors in the intensive care unit of a teaching and research hospital: A prospective cohort study. *Med Sci Monit* 2011;17:29-34.
5. Meric M, Willke A, Caglayan C, et al. Intensive care unit-acquired infections: incidence, risk factors and associated mortality in a Turkish university hospital *Jpn J Infect Dis* 2005;58:297-302.
6. Appelgren P, Hellström I, Weitzberg E, et al. Risk factors for nosocomial intensive care infection: a long-term prospective analysis. *Acta Anaesthesiol Scand* 2001;45:710-719.
7. Yilmaz GR, Cevik MA, Erdinc FS, et al. The risk factors for infections acquired by cerebral hemorrhage and cerebral infarct patients in a neurology intensive care unit in Turkey. *Jpn J Infect Dis*2007;60:87-91.
8. Vincent JL, Rello J, Marshall J, Silva E, Anzueto A, Martin CD, et al. International study of the prevalence and outcomes of infection in intensive care units. *JAMA* 2009; 302(21): 2323-2329.
9. Pradhan NP, Bhat SM, Ghadage DP. Nosocomial infections in the medical ICU: a retrospective study highlighting their prevalence, microbiological profile and impact on ICU stay and mortality. *J Assoc Physicians India* 2014;62:18-21.
10. Shalini S, Kranthi K, Gopalkrishna Bhat K. Microbiological profile of nosocomial infection in the intensive care unit. *J Clin Diagnostic Res* 2010; 4(5): 3109-3012.
11. Erb A, Sturmer T, Brenner H. Prevalence of antibiotic resistance in *Escherichia coli*: overview of geographical, temporal, and methodological variations. *Eur J Clin Microbiol Infect Dis* 2007; 26: 83-90.
12. Mukhopadhyay C, Bhargava A, Ayyagari A. Role of mechanical ventilation and development of multidrug resistant organisms in hospital acquired pneumonia. *Indian J Med Res.*2003;118:229–35.
13. Winarto. Prevalence of extended-spectrum -lactamases (ESBL)- bacteria of blood isolates in Dr. Kariadi Hospital Semarang 2004-2005. *Media Medika Indosiana* 2009; 43(5): 260-267.
14. Mehta A, Rosenthal VD, Mehta Y, Chakravarthy M, Todi SK, Sen N, et al. Device associated nosocomial infection rates in intensive care units of seven Indian cities: findings of the International Nosocomial Infection Control Consortium (INICC).
15. Hassandeh P, Motamedifar M, Hadi N. Prevalent bacterial infection in intensive care units of Shiraz University of Medical Science Teaching Hospital, Shiraz Iran. *Jpn J Infect Dis* 2009; 62: 249-253.
16. Chastre J, Fagon JY. Ventilator associated pneumonia. *Am J Respir Crit Care Med.*2002;165:867–903.
17. Gonlugur U, Bakici MZ, Akkurt I, Efeoglu T. Antibiotic susceptibility patterns among respiratory isolates of Gram negative bacilli in Turkish University Hospital. *BMC Microbiol.* 2004;4:32–4.
18. Ponce de LeónRosales SP, MolinarRamos F, DomínguezCherit G, et al. Prevalence of infections in intensive care units in Mexico: a multicenterstudy. *Crit Care Med* 2000; 28:13161321
19. Esen S, Leblebicioglu H. Prevalence of nosocomial infections at Intensive care units in Turkey: a multicentre 1-day point prevalence study. *Scan J Infect Dis* 2004;36:144-148.
20. Blot S. Limiting the attributable mortality of nosocomial infection and multidrug

- resistance in intensive care units. Clin Microbiol Infect 2008;14:5-13.
21. Brusselaers N, Vogelaers D, Blot S. The rising problem of antimicrobial resistance in the intensive care unit. Ann Intensive Care 2011;1:47.
 22. Katsaragakis S, Markogiannakis H, Samara E, et al. Predictors of mortality of Acinetobacter baumannii infections: A 2-year prospective study in a Greek surgical intensive care unit. Am J Infect Control 2010;38:631-635.
 23. Craven DE, Kunches LM, Lichtenberg DA, et al. Nosocomial and fatality in medical surgical intensive care unit patients. Arch Intern Med 1988;148:1161-1168.
 24. Mahzounieh M, Khoshnood S, Ebrahimi A, Habibian S, Yaghoubian M. Detection of Antiseptic-Resistance Genes in Pseudomonas and Acinetobacter spp. Isolated From Burn Patients. Jundishapur. J Nat Pharm Prod. 2014;9(2):e15402.
 25. Navaneeth BV, Belwadi MR. Antibiotic resistance among gram negative bacteria of lower respiratory tract secretion in hospitalized patients. Indian J Chest Dis Allied Sci. 2002;44:173-6.
 26. Datta P, Rani H, Chauhan R, Gombar S, Chander J. Health-care-associated infections: Risk factors and epidemiology from an intensive care unit in Northern India. Indian J Anaesth. 2014;58(1):30-5.
 27. Richards MJ, Edwards JR, Culver DH, et al. Nosocomial infections in medical intensive care units in the United States. National Nosocomial Infections Surveillance System. Crit Care Med 1999;27:887-892.
 28. Chelazzi C, Pettini E, Villa G, De Gaudio AR. Epidemiology, associated factors and outcomes of ICU-acquired infections caused by Gram-negative bacteria in critically ill patients: an observational, retrospective study. BMC Anesthesiol. 2015;15:125.
 29. Kumari HB, Nagarathna S, Chandramuki A. Antimicrobial resistance pattern among aerobic gram-negative bacilli of lower respiratory tract specimens of intensive care unit patients in a neurocentre. Indian J Chest Dis Allied Sci 2007; 49: 19-22.
 30. Falagas ME, Karveli EA, Siempos II, Vardakas KZ. Acinetobacter infections: a growing threat for critically ill patients. Epidemiol Infect. 2008 ;136(8):1009-19.